

## RESEARCH ARTICLE

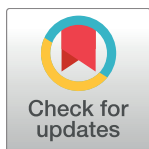
# Triploid atlantic salmon (*Salmo salar* L.) post-smolts accumulate prevalence more slowly than diploid salmon following bath challenge with salmonid alphavirus subtype 3

Lindsey J. Moore<sup>1‡</sup>, Tom Ole Nilsen<sup>2</sup>, Jiraporn Jarungsriapisit<sup>1,3</sup>, Per Gunnar Fjellidal<sup>4</sup>, Sigurd O. Stefansson<sup>3</sup>, Geir Lasse Taranger<sup>1</sup>, Sonal Patel<sup>1\*</sup>

**1** Institute of Marine Research, Bergen, Norway, **2** Uni Research Environment, Bergen, Norway, **3** Department of Biology, University of Bergen, Bergen, Norway, **4** Institute of Marine Research, Matre, Norway

‡ Current Address: Department of Biology, University of Bergen, Bergen, Norway

\* [sonal.patel@imr.no](mailto:sonal.patel@imr.no)



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## Abstract

Triploid Atlantic salmon (*Salmo salar* L.) may play an important role in the sustainable expansion of the Norwegian aquaculture industry. Therefore, the susceptibility of triploid salmon to common infections such as salmonid alphavirus (SAV), the causative agent of pancreas disease (PD), requires investigation. In this study, shortly after seawater transfer, diploid and triploid post-smolts were exposed to SAV type 3 (SAV3) using a bath challenge model where the infectious dose was 48 TCID<sub>50</sub> ml<sup>-1</sup> of tank water. Copy number analysis of SAV3 RNA in heart tissue showed that there was no difference in viral loads between the diploids and triploids. Prevalence reached 100% by the end of the 35-day experimental period in both infected groups. However, prevalence accumulated more slowly in the triploid group reaching 19% and 56% at 14 and 21 days post exposure (dpe) respectively. Whereas prevalence in the diploid group was 82% and 100% at the same time points indicating some differences between diploid and triploid fish. Both heart and pancreas from infected groups at 14 dpe showed typical histopathological changes associated with pancreas disease. Observation of this slower accumulation of prevalence following a natural infection route was possible due to the early sampling points and the exposure to a relatively low dose of virus. The triploid salmon in this study were not more susceptible to SAV3 than diploid salmon indicating that they could be used commercially to reduce the environmental impact of escaped farmed fish interbreeding with wild salmon. This is important information regarding the future use of triploid fish in large scale aquaculture where SAV3 is a financial threat to increased production.

**Competing interests:** The authors have declared that no competing interests exist.

**Abbreviations:** ADP, adenine diphosphate; CHH1, Chum salmon heart cells 1; cRNA, synthetic RNA; dpe, days post exposure; FCS, foetal calf serum; NKA, sodium/potassium ATPase activity; L:D, light:dark; PD, pancreas disease; SAV3, salmonid alphavirus type 3; TCID<sub>50</sub>, 50% tissue culture infectious dose.

## Introduction

Sustainable growth of the aquaculture industry in Norway and indeed globally is dependent on solving both disease and environmental issues. In the aquaculture of Atlantic salmon in Norway, major concerns are an increasing environmental footprint and challenges associated with salmon lice infestations and many viral diseases.

One of the environmental concerns is the Atlantic salmon escaping from marine net pens and interbreeding with wild salmon populations and it has been reported that the degree of introgression is largely dependent on the density of aquaculture sites nearby wild salmon rivers [1]. The use of sterile, triploid Atlantic salmon [2] in culture would avoid farmed, escaped salmon from reproducing in natural wild salmonid habitats. Triploid salmon, although phenotypically indistinguishable from diploids, have shown physiological differences requiring optimization of rearing conditions compared to diploids. This has manifested itself as a relative intolerance to warmer seawater temperatures compared to diploid salmon [3, 4]; the bulk production of triploid salmon takes place in the cooler northern Norwegian waters. Another challenge when rearing triploid salmon has been the optimization of feed composition to overcome skeletal deformities and cataracts that were more common in triploid salmon [5, 6].

Another concern in salmon aquaculture is viral diseases. One of the viral agents, salmonid alphavirus (SAV) causes pancreas disease (PD) in farmed salmonids in northern Europe during the seawater phase causing financial losses from both mortality and reduced filet quality [7]. SAV infection causes inflammation and cellular degeneration in target organs, beginning in exocrine pancreas and followed by heart and then skeletal muscle. PD is a debilitating disease causing fish to lose appetite and although many fish recover they fail to thrive and achieve good market weight. Mortality associated with PD outbreaks can vary between 5 and 90% due to several factors such as virulence of SAV subtype and isolate, husbandry, handling stress and may also depend on the robustness of the fish [8–12].

The susceptibility to diseases commonly encountered in aquaculture needs to be thoroughly evaluated to assess the suitability of triploid fish for commercial use. Comparison of diploid and triploid salmon has been carried out with regard to salmon lice, *Renibacterium salmoninarum* infection and *Aeromonas salmonicida* challenge following vaccination [13–15]. Thus, the susceptibility of triploid salmon to SAV3, and the subsequent development of PD in triploid fish is important information for the future development of the aquaculture industry. In order to mimic a natural route of infection, an established bath challenge model for use with Atlantic salmon post-smolts [16] was chosen for this study. It has been shown that the time taken from exposure to reach a prevalence of 100% within a population is dependent upon the exposure dose [17]. Therefore, diploid and triploid Atlantic salmon post-smolts were bath-challenged with SAV3 to evaluate their susceptibility to infection with a relatively low dose. The strength of this study is the use of a bath challenge model, early sampling time-points and a relatively low dose of virus that was instrumental in unravelling the observed differences in susceptibility to SAV3 in diploid and triploid salmon.

## Materials and methods

### Fish

AquaGen AS supplied diploid and triploid eyed eggs. To achieve triploidisation, immediately after fertilisation, the eggs were split into two equal batches, one batch for diploid and the other for the production of triploids. Triploidisation was performed according to Johnstone and Stet [18]. Briefly, shortly after fertilization the eggs were treated with a hydrostatic pressure shock of 655 bar, which prevents the expulsion of the second polar body and creates triploid

embryos of both sexes. In March 2015, Atlantic salmon eyed eggs both diploid and triploid (strain, AquaGen® Atlantic QTL-innOva® IPN/PD) were purchased from AquaGen AS and transferred to Matre Research Station, Institute of Marine Research. The eyed eggs were incubated at 6°C, until hatching. The diploid and triploid fry were kept in separate tanks. The salmon fry were first fed with commercial feed, with no extra nutrient additions for triploids (Skretting AS). The photoperiod was kept at L:D 24:0 until November 2015, when it was changed to L:D 12:12. On the 15th January 2016, the photoperiod was switched back to L:D 24:0 to induce parr-smolt transformation which prepares smolts for life in seawater. In mid-February 2016, diploid and triploid Atlantic salmon smolts were transferred in fresh water to the experimental facilities at the Industrial and Aquatic Laboratory (ILAB), Bergen, Norway. They were kept at 12°C, fed according to appetite with commercial feed, containing no extra nutrient additions for triploids (Skretting AS), and starved for 24 hours before sampling and handling. Fish were anaesthetized using 10 mg/L Metomidate and 60 mg/L Benzocaine before handling and euthanized with 10 mg/L Metomidate and 160 mg/L of Benzocaine before sampling. On the day the experiment started the water temperature was increased to 14°C and maintained at this temperature for the duration of the experiment.

## Virus

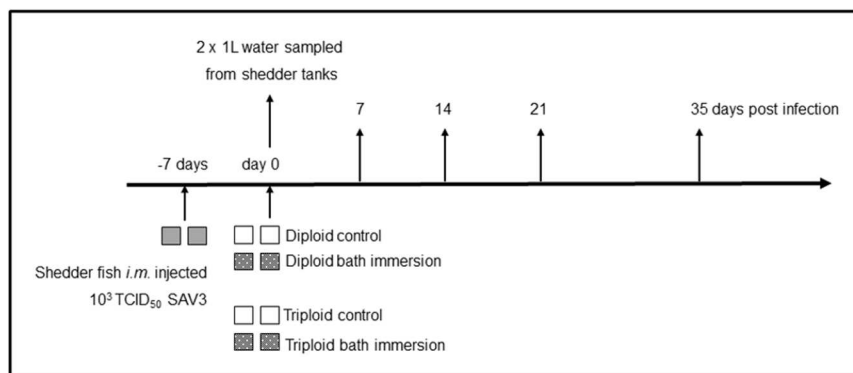
SAV3 strain H10 [19] was grown in CHH1 cells in L-15 medium supplemented with 2% FCS (foetal calf serum) and gentamycin (500µg/ml) at 15°C. The amount of infectious SAV3 in the cell supernatant was  $5 \times 10^6$  ml<sup>-1</sup> TCID<sub>50</sub> quantified by end-point dilution.

## Experimental procedures

Diploid and triploid smolts that were experimental fish were transferred to seawater (30 ‰) 4 days prior to the start of the experiment and maintained at 12°C. One hundred and thirty additional diploid salmon used as shedder fish were treated similarly. Diploid shedder fish were intramuscularly (*i.m.*) injected 1 week prior to the experimental bath challenge (-7 days) with  $1.3 \times 10^3$  TCID<sub>50</sub> SAV3 diluted in L-15 cell culture medium and kept in two 150 L tanks (Fig 1). The virus shed by these fish constituted the SAV3 used for the subsequent challenge of diploid and triploid fish. The day the experiment started the water flow was stopped for one hour in the shedder tanks and the shedder fish were removed and euthanized. The tank water from the shedder tanks was pooled and diluted, 1:4. One hundred liters of this diluted seawater, containing infectious virus (shed by the shedder fish) was added into each of the four 150 L tanks where the fish were exposed to SAV3. Two of these four tanks containing SAV3 were populated with 55 diploid fish per tank and the other two tanks with 55 triploid fish per tank. Four control tanks containing the same volume of water (100 L) were populated similarly with 55 diploid or triploid fish to act as non-infected control groups. The exposure to SAV3 was carried out for 6 hours, before the flow was re-started in all tanks according to Jarungsriapisit *et al.* [16]. The design of this experiment was approved by the Norwegian Animal Research Authority (ID: 8413).

## Sampling

Seawater (2 x 1 L) was sampled from each shedder tank, the day the experiment started after the flow had been stopped for one hour (Fig 1). Additional water samples were taken from the four tanks of diluted shedder water. Water samples were vacuum filtered through electropositive Zeta Plus® Virosoorb® 1 MDS filters (Cuno Inc., USA) using Millipore® sintered glass filters and funnels [20]. The virus was eluted from these filters in 1.2 mL of L-15 medium containing 10% FCS, thereby concentrating a litre of tank water into 1.2 mL according to



**Fig 1. Experimental set up.** The time-line line for the experiment showing all tanks and sampling time-points. At the start of the experiment, the average weight of the fish (random 10 fish within each group) was  $76.4 \pm 12.2$  g for diploids and  $71.6 \pm 19$  g for triploids.

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Jarungsriapisit *et al.* [17]. The virus in the eluant was quantified by measuring SAV3 RNA by qPCR (see below) and by TCID<sub>50</sub> end-point titration in order to determine the bath immersion dose. Eight post-smolts were sampled from each tank at 7, 14, 21 and 35 days post exposure (dpe), and their weights and fork-lengths recorded for condition factor calculation using the following formula:

$$CF = [\text{fish weight (g)} \times 100] / [\text{fish length (cm)}]^3$$

Heart samples from all sampled fish were flash frozen in liquid nitrogen and stored at  $-80^\circ\text{C}$  until analysis. Heart and pancreas tissues were sampled for histology from the same 8 fish at 14 dpe when typical PD pathology is most commonly observed in both heart and pancreas and fixed in 10% neutral buffered formalin. Adipose fins were collected and stored in ethanol from all sampled triploid fish to verify triploidy.

## Total RNA preparation

Heart tissue samples (half the heart) were homogenized in 1 ml TRIzol® using a Precellys 24 instrument and 1.4 mm diameter Zirconium oxide beads. After the homogenization and chloroform treatment, 450 µl of supernatants were used further for RNA isolation using a Purelink total RNA extraction kit, in an iPrep machine (Life Technologies). RNA from filtered/concentrated seawater eluants was isolated by mixing 100 µl eluant with 350 µl lysis buffer, and further isolation was carried out by Purelink total RNA extraction kit (Life Technologies). RNA was eluted in 50 µl of the propriety buffer. RNA concentration and quality was estimated using a Nanodrop 1000 ND. Ten percent of the samples were checked for integrity on a Bioanalyser (Agilent Instruments) resulting in RINs  $\geq 8$ .

## cDNA and RT-qPCR for viral copy number quantification

SAV3 RNA from heart tissue was quantified with a modified one-step nsP-1 assay (Ag-Path, Ambion) [21] with a sense probe, using 200 ng total RNA in a total reaction volume of 10 µl. Approximately twenty percent of heart RNA samples randomly selected from all groups and time-points were qualitatively verified by measuring the transcription of elongation factor 1A [22]

SAV3 RNA isolated from filtered/concentrated water samples was quantified using a two-step assay and together with the TCID<sub>50</sub> result constituted the virus dose the fish were exposed

to. cDNA was transcribed from 2  $\mu$ l total RNA in a 10  $\mu$ l reaction using SuperScript™ VILO™ (Invitrogen) as described in the manufacturer's instructions. qPCR was run in triplicate in 96 well plates using TaqMan® Fast Universal Master Mix (Applied Biosystems®) and an Applied Biosystems 7900H Fast sequence detection analyser. A 10  $\mu$ l reaction volume contained; 2  $\mu$ l cDNA (diluted 1:10), 5  $\mu$ l 2 x master mix, 900nM of each primer and 250nM of FAM-labelled probe. The running conditions were as recommended by the manufacturer.

SAV3 RNA was quantified using a standard curve produced using a 10 x dilution series of synthetic SAV3 RNA (576 bps, cRNA) containing  $10^1$  to  $10^7$  copies of SAV RNA [17]. These standards were analysed on each plate when quantifying copy numbers from heart RNA in the one-step assay. The same dilutions were reverse transcribed (as above) and run in duplicate for the quantification of copy numbers in the two-step assay for water samples.

## Histology

Formalin fixed tissues were paraffin embedded and 3 $\mu$ m sections Haematoxylin-Erythrosin-Saffron (HES) stained before visualizing under a Leica DMRBE light microscope (Leica Microsystems, Germany). Photographs were taken using Spotflex camera model nr 15.2 64 Mp Shifting pixel (Diagnostic instruments Inc, USA) and processed with Image J software.

## ATPase analysis

Smolt status was analysed by measurement of  $\text{Na}^+/\text{K}^+$  ATPase activity (NKA) in gill tissue 2 weeks before the start of the experiment and before the fish were transferred to the experimental facility. Analysis was carried out as described previously [23] and results expressed as  $\mu\text{mol ADP mg protein}^{-1} \text{ h}^{-1}$ .

## Triploid determination

Before transfer to the experimental facility ploidy was verified by measuring erythrocyte diameter on blood smears on 10 triploids [24]. For fish sampled during the experiment genomic DNA was extracted (Qiagen, DNeasy) from adipose fins collected from all triploid fish. Triploid status was verified using multi-loci microsatellite genetic profiling; the presence of three alleles at multiple loci [25], and any fish not triploid at all loci investigated were excluded from further analysis.

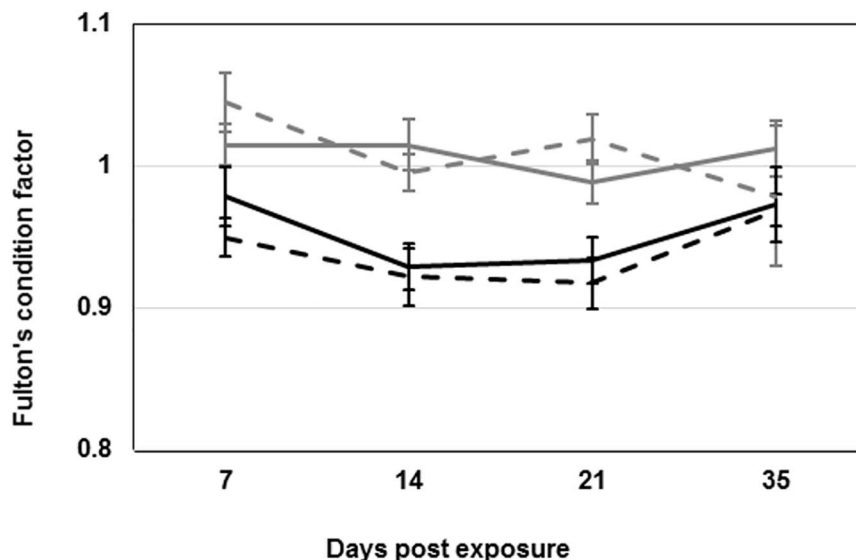
## Data analysis

SDS software version 2.4.1 automatically calculated copy numbers of the nsP-1 transcript from the Ct values of a standard curve run on the same plate. These copy numbers were analysed using Statistica version 12.7. Condition factors between infected and control fish within diploid and triploid groups at all time-points were compared using a t test (MS Excel 2016).

## Results

### Fish

All fish within the triploid group were triploid when tested before transfer to ILAB (erythrocyte size on blood smears). Of the 74 experimental fish tested (multiple loci microsatellite mapping) one was diploid and one sample was contaminated and these 2 fish were excluded from further analysis. Condition factors calculated from weights and lengths of all fish remained stable and were not significantly different within the diploid and triploid salmon when compared to their respective non-infected controls throughout the experiment (Fig 2).



**Fig 2. Condition factor.** Fulton's condition factor calculated by:  $100 \times \text{weight (g)} / [\text{fork length (cm)}]^3$  for all fish sampled at all time-points. Lines represent the average  $\pm$  SEM per group. Black lines represent triploid groups and grey lines the diploid groups. Dashed lines represent control groups.

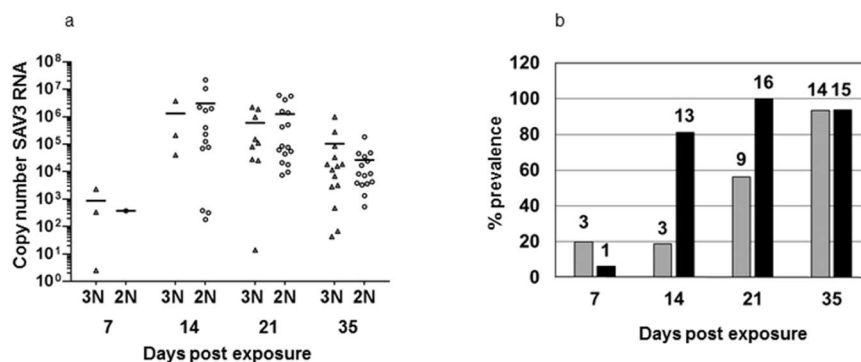
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## Bath immersion dose

On the day of exposure, water from the shedder tanks contained an average of 160 copies of SAV3 RNA per litre and the TCID<sub>50</sub> assay revealed 48 TCID<sub>50</sub> SAV3 per litre. The shedder water was diluted 1:4 and used for exposure to potentially infect the diploid and triploid salmon. Water sampled after the 1:4 dilution produced unreliable results in the copy number analysis and, therefore, the result for the undiluted water was used and divided by 4 (the dilution factor).

## PD status

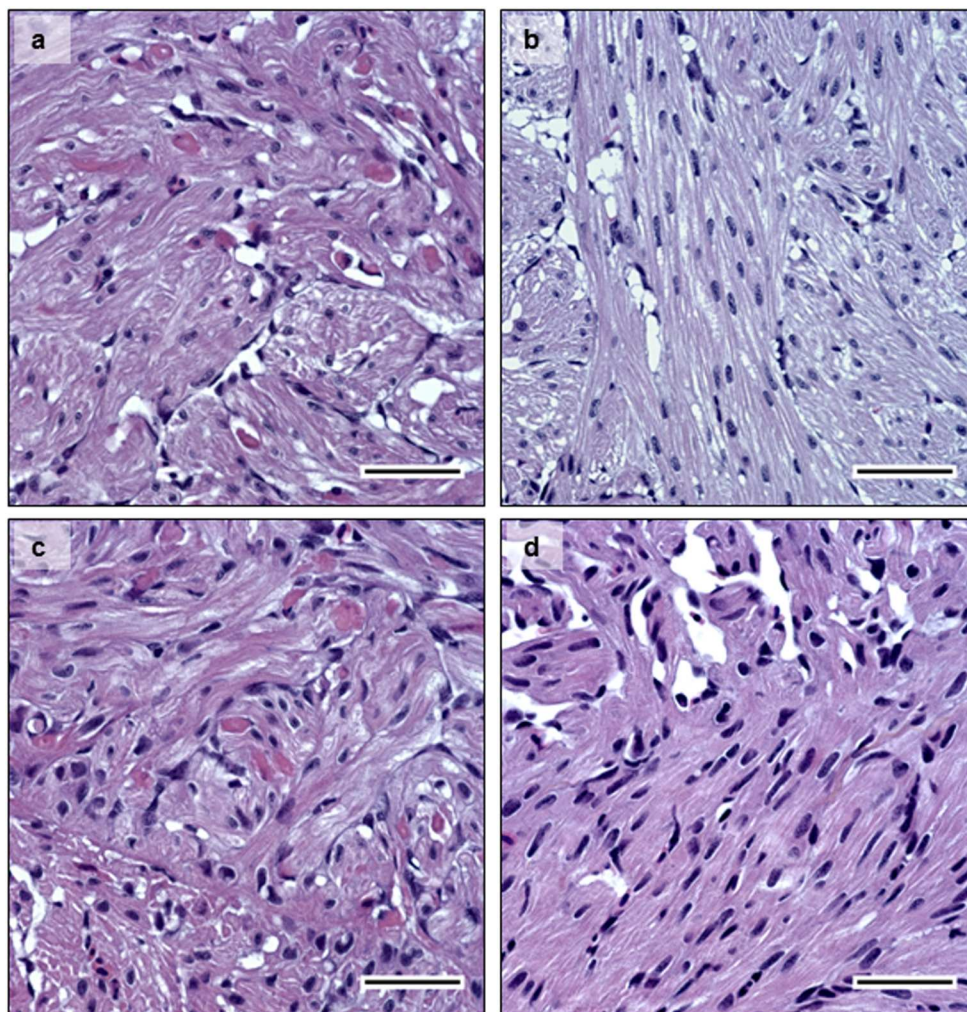
SAV3 status was determined by copy number analysis of SAV3 RNA in total RNA from heart tissue (Fig 3A). Copy number analysis showed a 100% prevalence in the diploid group by 21



**Fig 3. SAV3 Status.** 3a. Copy numbers of SAV3 RNA (viral load) of all individuals in triploid ( $\Delta$ , 3N) and diploid (O, 2N) groups at all time-points. 3b. Percentage positive fish for SAV3 RNA in the triploid group (grey bars) and diploid group (black bars) at all time-points. Numbers above the columns are the number of positive fish ( $n = 16$  except in the triploid group at 7 and 35 dpe where  $n = 15$ ).

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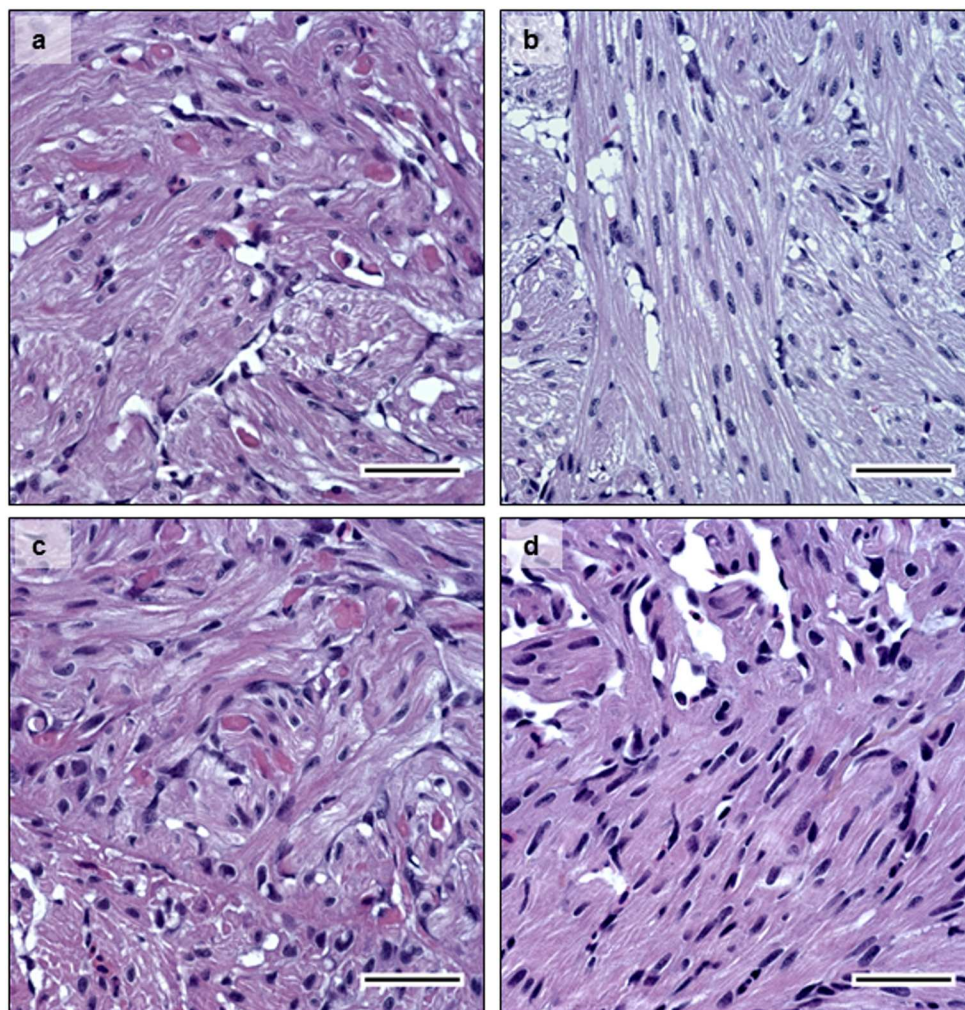
**Fig 4. Histology.** HES staining of pancreas tissue sections at 14 dpe. **4a-** infected diploid, **4c-** infected triploid, showing cell degeneration. **4b-** non-infected diploid; and **4d-** non-infected triploid. Infected fish showed loss of exocrine pancreatic cells and immune cell infiltration, while non-infected controls show normal histology in pancreas. Bar = 50μm.

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dpe, whereas the prevalence in the triploid group rose more slowly reaching 93.3% by the end of the experiment (35 dpe) (Fig 3B). Although slower to reach a maximum prevalence the triploid group showed similar viral loads (copy numbers of SAV3 RNA) to the diploid group at all time-points (Fig 3A). There was no significant difference in the viral loads between the diploid and the triploid groups ( $p \geq 0.05$ ) at any of the time-points.

Additionally, PD status was confirmed histologically at 14 dpe when pathology is usually observed in both heart and pancreas. PD-associated lesions such as degeneration of exocrine pancreatic cells (Fig 4A–diploid, and 4B–triploid), and degeneration of myocardial cells in the heart were observed (Fig 5A–diploid, and 5B–triploid). Loss of exocrine pancreatic cells and infiltration by inflammatory cells was observed in pancreas tissue at 14 dpe in both diploid and triploid fish (Fig 4A and 4B respectively). The degree of histopathological changes in the infected individuals was independent of whether the fish were diploid or triploid. None of the control fish examined showed any PD-associated histopathological changes in the pancreas (Fig 4C and 4D), or in the heart (Fig 5C and 5D).





**Fig 5. Histology.** HES staining of heart tissue sections at 14 dpe. **5a-** infected diploid, **5b-** infected triploid, showing myocardial cell degeneration. **5c-** non-infected diploid; and **5d-** non-infected triploid, show normal histology in heart tissue sections. Bar = 50μm.

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### Na<sup>+</sup>/K<sup>+</sup> ATPase status

High gill NKA activities in all diploid and triploid fish tested before the start of the experiment ( $n = 10$ ) showed that these groups of fish were ready for seawater transfer. Diploid fish had an average of  $14.6 \mu\text{mol ADP mg protein}^{-1} \text{ h}^{-1}$  (range: 11.0 to 20.0) and triploid fish an average of  $16.3 \mu\text{mol ADP mg protein}^{-1} \text{ h}^{-1}$  (range: 12.3 to 21.7).

### Discussion

At 35 dpe 93% of both the diploid and the triploid SAV3 exposed fish were positive for SAV3 RNA. In addition, histological examination showed the development of PD-associated lesions in these groups. Prior to seawater transfer diploid and triploid smolts showed gill NKA activity commensurate with good quality smolts and good seawater tolerance. The aquaculture industry aims to transfer only good quality smolts to sea cages so that the fish will be robust and capable of fighting pathogens. However, it has been reported that even though salmon smolts are within the smolt window when transferred to seawater they are more susceptible to SAV3



during the first few weeks [16]. In marine sea cages, the dose of viral agents that salmon would be exposed to is anticipated to be very low. By choosing both a relatively low dose and smolts recently transferred to seawater this experiment was sufficiently sensitive to ensure that at least a few individuals would be infected, and the difference in the accumulation of prevalence between diploid and triploid salmon could be observed. Since both diploid and triploid salmon in this study were raised from the same batch of eggs, had comparable NKA activities before seawater transfer, and were exposed to virus simultaneously the differences in their accumulation of prevalence to SAV3 could be attributed to their difference in ploidy. The condition factor was not affected by SAV3 infection in the five weeks experiment in this study. However, a reduction in condition factor has been reported in a similar experiment with SAV3 and Atlantic salmon post-smolts during a longer 12 weeks experiment [9].

The exposure to SAV3 was estimated by both nsP-1 (SAV3 RNA) copy number analysis and by end-point dilution of infectious virus ( $TCID_{50} L^{-1}$  in the tank water). Although the numbers of virus per litre measured by  $TCID_{50}$  is considerably less than that measured by the copy number analysis, it represents a live infectious dose and not SAV3 RNA, and thus the results from the two methods are not directly comparable. Since prevalence approached 100% in all infected tanks, the exposure dose was clearly sufficient to cause infection.

Previous studies have failed to demonstrate major differences in disease resistance between diploid and triploid Atlantic salmon. An earlier study involving all female diploid and triploid groups of salmon was unable to find any differences in the susceptibility to bacterial kidney disease due to ploidy differences [13]. Triploid Chinook salmon (*Oncorhynchus tshawytscha*) showed an increased mortality following *Vibrio anguillarum* challenge, but only under stressful conditions [14]. Diploid and triploid siblings performed equally well when vaccinated and then challenged with *Aeromonas salmonicida* although there were differences in their white cell numbers and respiratory burst activity [15]. Finally, the susceptibility to sea lice infestation in triploid Atlantic salmon, investigated at different life stages both in Scotland and in Norway, concluded that triploid salmon showed no increased susceptibility to lice infestation compared to diploids [26].

The fish used in this study were from the same batch of AquaGen® QTLinnOva® IPN/PD strain that possess an increased resistance to viral diseases. They were infected using an experimental model that attempts to mimic the conditions at aquaculture sites, but both the exposure time and dose were likely higher than experienced during a field infection, thus 100% prevalence was attained rather rapidly compared to a field outbreak where prevalence increases more gradually within a sea-cage.

These results are from an experimental infection using relatively small groups of fish and a single isolate of SAV3. It could be of interest to investigate the effects of varying parameters such as other virus isolates or sub-types, size and physiological status of fish on the susceptibility of SAV3. Differences in mortality between isolates has been documented [9]. The temperature of 14°C used in this study is an optimum temperature for diploid salmon post-smolts [27, 28] of the weight used in this experiment, while triploid post-smolts have a lower temperature optimum [29]. In addition, all the fish in this study received standard feed for diploid fish although special nutrient formulations specifically for triploid fish to optimize their growth rates and reduce skeletal deformities have been recommended. However, sub-optimal temperature and sub-optimal feed had no detrimental effect on the maximal prevalence in the triploid group compared to the diploid group, and the prevalence accumulated more slowly in the triploid group.

It is not known why triploids during this experiment exhibited a slower accumulation of prevalence, but differences in the proportions of immune cells in the head kidney have been reported between triploid and diploid Atlantic salmon following vaccination [15, 30]. These

differences may however be compensating for the slightly larger cell size in triploid fish [31, 32], but could also indicate differences in the immune response in these fish groups. In conclusion, although no significant difference in the prevalence of SAV3 could be seen between triploids and diploids at the end of a 5 weeks experimental period, the virus appeared to spread more slowly through the triploid fish group. Under field conditions where even lower doses are likely this slower accumulation of prevalence could either delay the onset of PD or avoid an outbreak altogether. Such a scenario would be a significant advantage in the months before harvest when mortality from PD is financially the most damaging. Further investigation of possible differences when triploids and diploids are exposed to very low amounts of virus, more closely mimicking field conditions, would be interesting. The triploid group in this study showed no increased susceptibility or pathology when exposed to SAV3 virus indicating that their commercial use could reduce the environmental impact of escaped farmed fish interbreeding with wild salmon.

## Supporting information

**S1 Raw data.**  
(XLSX)

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## Author Contributions

**Conceptualization:** SS GLT SP.

**Data curation:** LJM TON SP.

**Formal analysis:** LJM SP.

**Funding acquisition:** SP.

**Investigation:** LJM JJ SP.

**Methodology:** LJM TON SS GLT PGF JJ SP.

**Project administration:** SP.

**Resources:** LJM PGF TON SP.

**Supervision:** SP.

**Validation:** LJM SP.

**Visualization:** LJM SP.

**Writing – original draft:** LJM SP.

**Writing – review & editing:** LJM TON, SS GLT PGF JJ SP.

## References

1. Glover KA, Pertoldi C, Besnier F, Wennevik V, Kent M, Skaala Ø. Atlantic salmon populations invaded by farmed escapees: quantifying genetic introgression with a Bayesian approach and SNPs. *BMC Genetics*. 2013; 14(1):1–19.
2. Benfey TJ, Dye HM, Solar II, Donaldson EM. The growth and reproductive endocrinology of adult triploid Pacific salmonids. *Fish physiology and biochemistry*. 1989; 6(2):113–20. <https://doi.org/10.1007/BF01875483> PMID: 24226976
3. Galbreath PF, Adams ND, Sherrill LW, Martin TH. Thermal Tolerance of Diploid Versus Triploid Rainbow Trout and Brook Trout Assessed by Time to Chronic Lethal Maximum. *Environmental Biology of Fishes*. 2006; 75(2):183–93.
4. Maxime V. The physiology of triploid fish: current knowledge and comparisons with diploid fish. *Fish and Fisheries*. 2008; 9(1):67–78.
5. Fjellidal P, Hansen T, Lock EJ, Wargelius A, Fraser T, Sambras F, et al. Increased dietary phosphorous prevents vertebral deformities in triploid Atlantic salmon (*Salmo salar* L.). *Aquaculture Nutrition*. 2016; 22(1):72–90.
6. Sambras F, Fjellidal P, Remø S, Hevrøy E, Nilsen T, Thorsen A, et al. Water temperature and dietary histidine affect cataract formation in Atlantic salmon (*Salmo salar* L.) diploid and triploid yearling smolt. *J Fish Dis*. 2017
7. Lerfall J, Larsson T, Birkeland S, Taksdal T, Dalgaard P, Afanasyev S, et al. Effect of pancreas disease (PD) on quality attributes of raw and smoked fillets of Atlantic salmon (*Salmo salar* L.). *Aquaculture*. 2012; 324:209–17.
8. Graham DA, Frost P, McLaughlin K, Rowley HM, Gabestad I, Gordon A, et al. A comparative study of marine salmonid alphavirus subtypes 1–6 using an experimental cohabitation challenge model. *J Fish Dis*. 2011; 34(4):273–86. <https://doi.org/10.1111/j.1365-2761.2010.01234.x> PMID: 21294751
9. Taksdal T, Bang Jensen B, Bockerman I, McLoughlin MF, Hjortaas MJ, Ramstad A, et al. Mortality and weight loss of Atlantic salmon, *Salmo salar* L., experimentally infected with salmonid alphavirus subtype 2 and subtype 3 isolates from Norway. *J Fish Dis*. 2014.
10. Kristoffersen A, Viljugrein H, Kongtorp R, Brun E, Jansen P. Risk factors for pancreas disease (PD) outbreaks in farmed Atlantic salmon and rainbow trout in Norway during 2003–2007. *Prev Vet Med*. 2009; 90.
11. Jansen MD, Taksdal T, Wasmuth MA, Gjerset B, Brun E, Olsen AB, et al. Salmonid alphavirus (SAV) and pancreas disease (PD) in Atlantic salmon, *Salmo salar* L., in freshwater and seawater sites in Norway from 2006 to 2008. *J Fish Dis*. 2010; 33(5):391–402. <https://doi.org/10.1111/j.1365-2761.2009.01131.x> PMID: 20158578
12. McLoughlin M, Graham D. Alphavirus infections in salmonids—a review. *J Fish Dis*. 2007; 30.
13. Bruno DW, Johnstone R. Susceptibility of diploid and triploid Atlantic salmon *Salmo salar* L., to challenge by *Renibacterium salmoninarum*. *Bull Eur Ass Fish Pathol*. 1990; 10(2):45–7.
14. Ching B, Jamieson S, Heath JW, Heath DD, Hubberstey A. Transcriptional differences between triploid and diploid Chinook salmon (*Oncorhynchus tshawytscha*) during live *Vibrio anguillarum* challenge. *Heredity*. 2010; 104(2):224–34. <https://doi.org/10.1038/hdy.2009.108> PMID: 19707232
15. Chalmers L, Thompson KD, Taylor JF, Black S, Migaud H, North B, et al. A comparison of the response of diploid and triploid Atlantic salmon (*Salmo salar*) siblings to a commercial furunculosis vaccine and subsequent experimental infection with *Aeromonas salmonicida*. *Fish Shellfish Immun*. 2016; 57:301–8.
16. Jarungsriapisit J, Moore LJ, Taranger GL, Nilsen TO, Morton HC, Fiksdal IU, et al. Atlantic salmon (*Salmo salar* L.) post-smolts challenged two or nine weeks after seawater-transfer show differences in their susceptibility to salmonid alphavirus subtype 3 (SAV3). *Virol J*. 2016; 13(1):1–14.
17. Jarungsriapisit J, Moore LJ, Mæhle S, Skår C, Einen AC, Fiksdal IU, et al. Relationship between viral dose and outcome of infection in Atlantic salmon, *Salmo salar* L., post-smolts bath-challenged with salmonid alphavirus subtype 3. *Vet Res*. 2016; 47(1):102. <https://doi.org/10.1186/s13567-016-0385-2> PMID: 27760562
18. Johnstone R, Stet RJM. The production of gynogenetic Atlantic salmon, *Salmo salar* L. Theoretical and Applied Genetics. 1995; 90(6):819–26. <https://doi.org/10.1007/BF00222017> PMID: 24172924
19. Xu C, Guo TC, Mutoloki S, Haugland O, Marjara IS, Evensen O. Alpha interferon and not gamma interferon inhibits salmonid alphavirus subtype 3 replication in vitro. *J Virol*. 2010; 84(17):8903–12. PubMed Central PMCID: PMC2919011. <https://doi.org/10.1128/JVI.00851-10> PMID: 20573808
20. Andersen L, Hodneland K, Nylund A. No influence of oxygen levels on pathogenesis and virus shedding in Salmonid alphavirus (SAV)-challenged Atlantic salmon (*Salmo salar* L.). *Virol J*. 2010; 7.

21. Jarungsriapisit J, Moore L. J., Maehle S. Skaar C., Einen A.C. B., Fiksdal I., Morton H. C., Stefansson S., Taranger G. L. and Patel S. Relationship between viral dose and outcome of infection in Atlantic salmon, *Salmo salar* L., post-smolts bath-challenged with salmonid alphavirus subtype 3. *J Vet Res.* 2016; 47:102–15.
22. Hodneland K, Endresen C. Sensitive and specific detection of Salmonid alphavirus using real-time PCR (TaqMan). *J Virol Methods.* 2006; 131(2):184–92. <https://doi.org/10.1016/j.jviromet.2005.08.012> PMID: 16202457
23. Lovoll M, Austbo L, Jorgensen JB, Rimstad E, Frost P. Transcription of reference genes used for quantitative RT-PCR in Atlantic salmon is affected by viral infection. *Vet Res.* 2011; 42:8–13. PubMed Central PMCID: PMC3031228. <https://doi.org/10.1186/1297-9716-42-8> PMID: 21314970
24. McCormick SD. Methods for nonlethal gill biopsy and measurement of Na<sup>+</sup>, K<sup>+</sup> -ATPase activity. *Can J Fish Aquat Sci.* 1993; 50.
25. Fjelldal PG, Hansen T. Vertebral deformities in triploid Atlantic salmon (*Salmo salar* L.) underyearling smolts. *Aquaculture.* 2010; 309(1–4):131–6.
26. Glover KA, Madhun AS, Dahle G, Sørvik AGE, Wennevik V, Skaala Ø, et al. The frequency of spontaneous triploidy in farmed Atlantic salmon produced in Norway during the period 2007–2014. *BMC Genetics.* 2015; 16(1):1–10.
27. Frenzl B, Migaud H, Fjelldal PG, Shinn AP, Taylor JF, Richards RH, et al. Triploid and diploid Atlantic salmon show similar susceptibility to infection with salmon lice *Lepeophtheirus salmonis*. *Pest management science.* 2014; 70(6):982–8. <https://doi.org/10.1002/ps.3639> PMID: 23983154
28. Handeland SO, Imsland AK, Stefansson SO. The effect of temperature and fish size on growth, feed intake, food conversion efficiency and stomach evacuation rate of Atlantic salmon post-smolts. *Aquaculture.* 2008; 283(1):36–42.
29. Sambras F. Solving bottlenecks in triploid Atlantic salmon production. Temperature, hypoxia and dietary effects on performance, cataracts and metabolism. PhD thesis. 2016; University of Bergen.
30. Hansen TJ, Olsen RE, Stien L, Oppedal F, Torgersen T, Breck O, et al. Effect of water oxygen level on performance of diploid and triploid Atlantic salmon post-smolts reared at high temperature. *Aquaculture.* 2015; 435:354–60.
31. Fraser TWK, Rønneseth A, Haugland GT, Fjelldal PG, Mayer I, Wergeland HI. The effect of triploidy and vaccination on neutrophils and B-cells in the peripheral blood and head kidney of 0+ and 1+ Atlantic salmon (*Salmo salar* L.) post-smolts. *Fish Shellfish Immun.* 2012; 33(1):60–6.
32. Budiño B, Cal RM, Piazzon MC, Lamas J. The activity of several components of the innate immune system in diploid and triploid turbot. *Comparative Biochemistry and Physiology Part A: Molecular & Integrative Physiology.* 2006; 145(1):108–13.